

#### ArrayTrack<sub>3,1,5</sub> Demonstration

National Center for Toxicological Research U.S. Food and Drug Administration 3900 NCTR Road, Jefferson, AR 72079





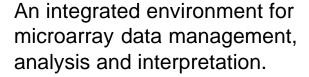
#### **ArrayTrack Overview**

#### Microarray DB

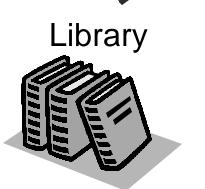




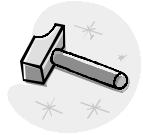




Using ArrayTrack, the user can apply analysis method from Tool to microarray DB and then get Information from the linked LIB for biological interpretation





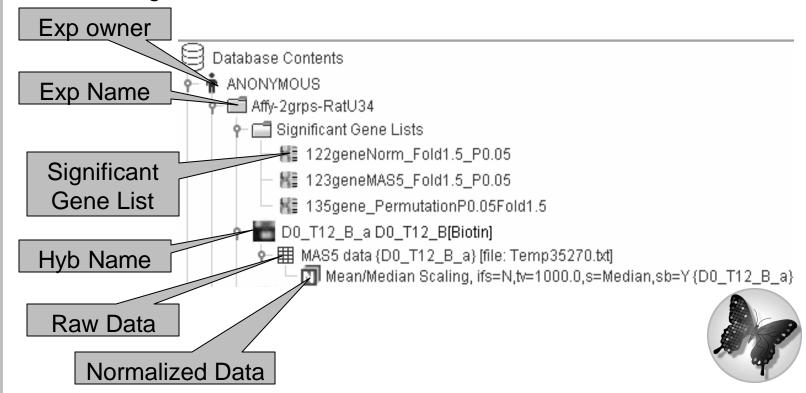






#### Microarray DB

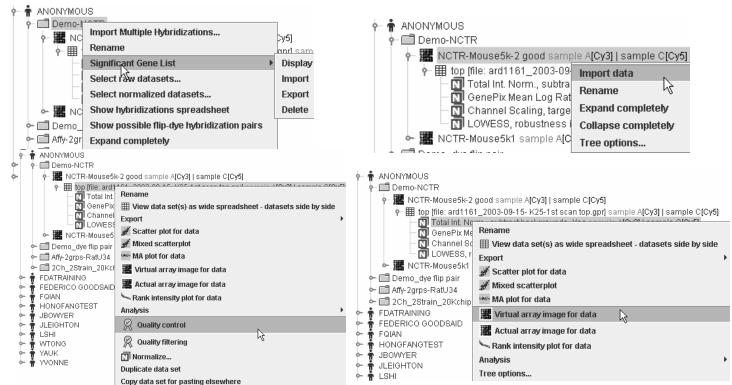
Data is organized as a hierarchical tree structure:





#### **Exploring DB**

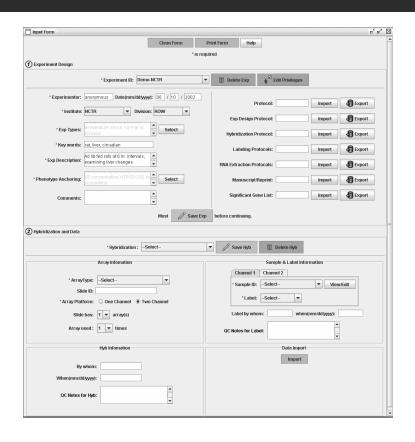
 Right-click exp, hybridization, raw data and normalized data to access various functions







#### **Importing Data**



- 1. create experiment
- create hybridization
- create array type (if not existed in AT Chip Lib)
- 4. create sample

MicroarrayDB stores DNA microarray gene expression experimental data together with essential annotation information about an experiment, its protocol and the samples.

Support data from one-channel and two-channel.



#### **Searching Libraries**

#### Library







🌉 Protein Library

**ini** IPI Library

💿 Orthologene Library

🐫 GOFFA Library

Chip Library

🌉 Toxicant Library

🔤 EDKB Library

There are nine libraries in ArrayTrack. All the libraries are interlinked.

The libraries integrate gene, protein, pathway and other data allowing data interrogation and mining of data across data types.

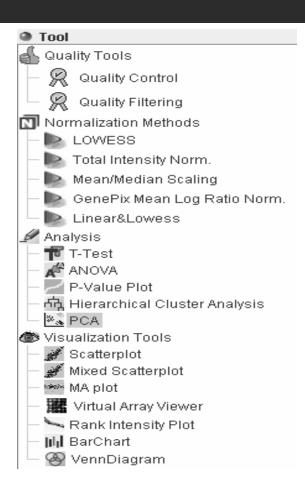
Several ways to activate these libraries

- -From library panel
- -From the Library pull-down menu
- -From the results of analysis (e.g. T-test)





### **Analysis Tools**



#### Tools:

#### Quality Tools

Provides various visual plots and numerical parameters for measuring the quality of a hyb, and filtering the unwanted spots.

#### Normalization Methods

Correct systematic variations in microarray data introduced by experimental factors

#### Analysis

Provides different methods to evaluate the microarray data

#### Visualization Tools

Provides a direct view to identify abnormalities within data.





#### Microarray Data Processing

Importing data

Loading array data into AT

Normalization

Ensure cross-chip comparison

Gene Selection

Identify a list of significant genes

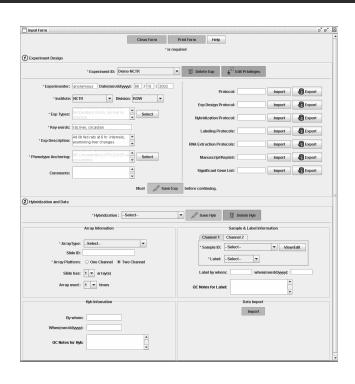
Interpretation

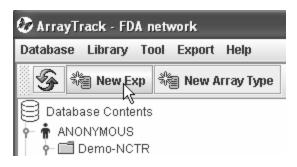
Interpret data using pathways and GO





#### **Importing Data**





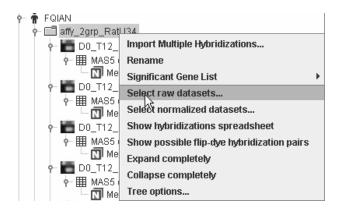
- 1. create experiment
- 2. create hybridization
- create array type (if not existed in AT Chip LIB)
- 4. create sample





#### Normalization

- To remove systematic variations across chips and ensure a valid cross-chip comparison
  - 1. Right-click an experiment
  - 2. Select "raw datasets..."



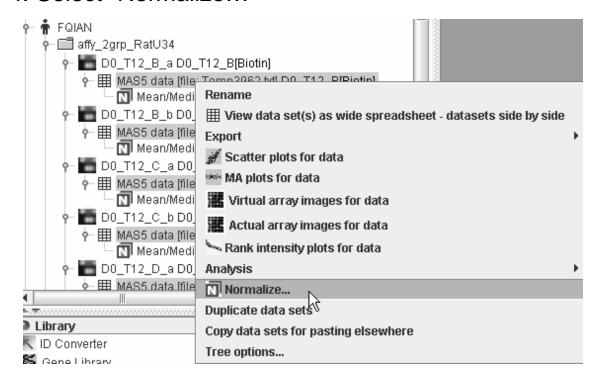






### Normalization (-continued)

- 3. Right-click any highlighted raw data
- 4. Select "Normalize..."

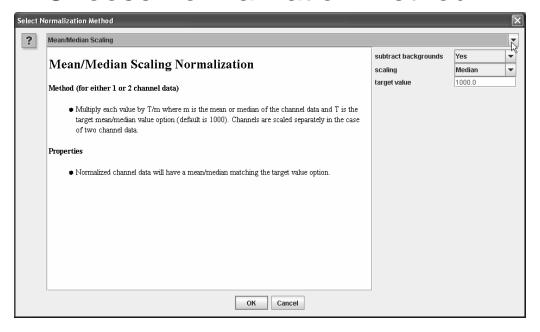






## Normalization (-continued)

Choose normalization method



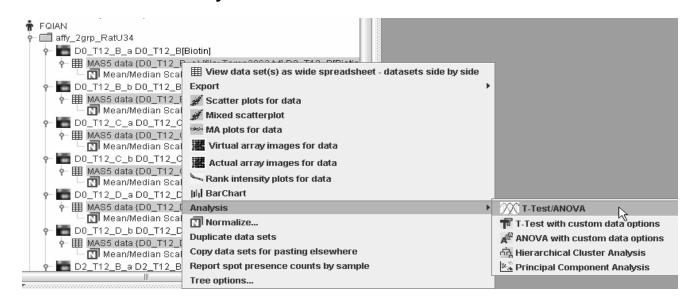
- •For Affy data, choose
- "Mean/Median Scaling"
  •For 2-channel data, the default method is "Lowess"





#### **Gene Selection**

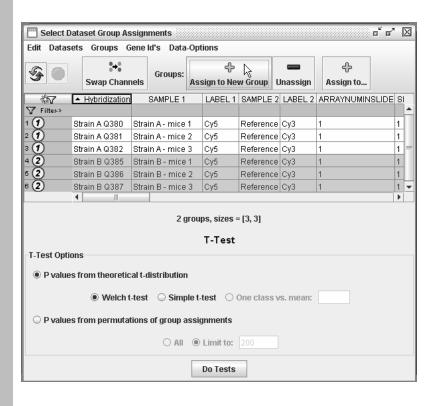
- Highlight and Right-click the experiment.
- Select "Normalized datasets..." and click "OK"
- Right-click the highlighted normalized data
- Choose "Analysis->T-test"







## **Gene Selection (-continued)**



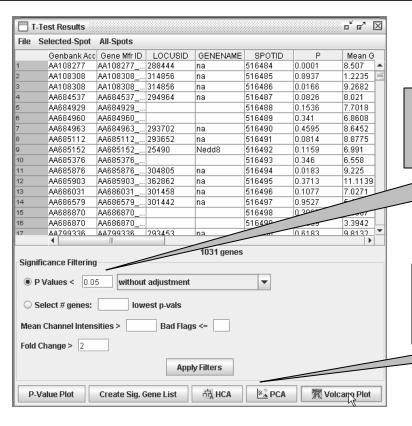
Assign the data into 2 groups

- -different dose
- -different time
- -or different animal





### **Gene Selection (-continued)**



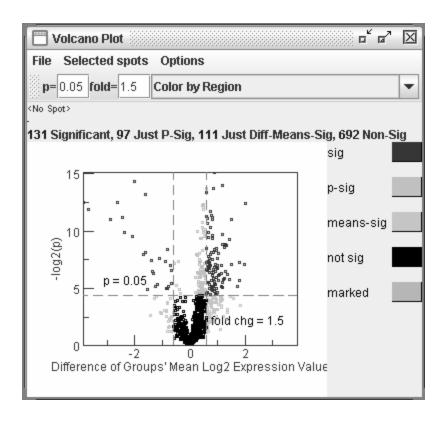
Parameters used to filter the genes to get significant gene list

Access other analysis methods like HCA, PCA, Volcano plot, etc.





## **Gene Selection (-continued)**



Volcano Plot



#### **Significant Gene List**

- Create
- Display
- Import
- Export
- Delete

### Significant Gene List(-cont)

Create significant gene list From T-test/Anova result T-Test Results All-Spots Advanced File Selected-Spot set filter criteria Create significant gene list... NM 01347 49926 6.4884 2,7783 13.0868 Hierarchical Clustering Analysis AY0655 46938 -2.0418 -3.79553.3721 Principal Component Analysis 51257 -0.7993 0.5981 e.g. P-value, fold change 1.3012 Library searches 🗲 Gene Library 0.0397 Select in all open viewers ₩. Pathway Library 3.2184 to get significant gene list U86105 U86105\_1 18113 Protein Library -0.6934 0.2812 AK018313 AK018313\_1 0.0007 0.242 1.1507 NM\_008865 NM\_00886... 18776 159805 0.0008 -0.8595 1.8513 NM\_019486 NM\_01948... 56015 0.2548 1.2377 NM\_008319 NM\_00831... 15898 145874 0.0009 -0.1588 -0.4882 1.2564 NM\_021447 NM\_02144... 58522 0.001 0.1626 1.1867 Rnf30 149205 0.4095 NM 015780 NM 01578.. 0.2485 Cfhl1 148342 0.0011 1.2908 3.2995 D43759 D43759\_1 14161 156273 0.0011 5.9168 4.6198 2.4571 NM 010419 NM 01041... 15208 157161 0.0011 1.1625 AF331708 AF331708 1 230103 0.2314 **Create significant gene list** AR041350 AR041350 1 12830 158418 0.0012 1.8511 20160 genes Significance Filtering without adjustment P Values <</p> Target False Discovery Rate (FDR): Filter criteria Select # genes with lowest p-vals: Bad Flags <= Mean Channel Intensities > Fold Change > Apply Filters P-Value Plot Create Sig. Gene List Volcano Plot

## Significant Gene List(-cont)

Display/Import/Export/Delete Significant Gene List

Right-click the experiment name

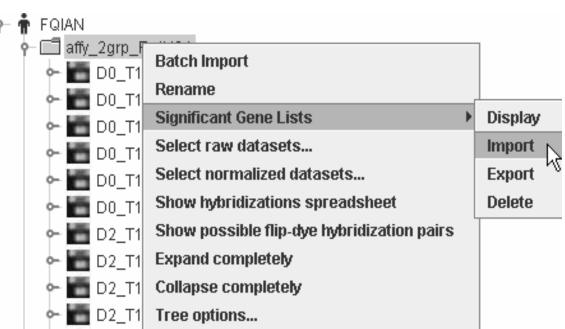
select Significant Gene Lists

select Display

**Import** 

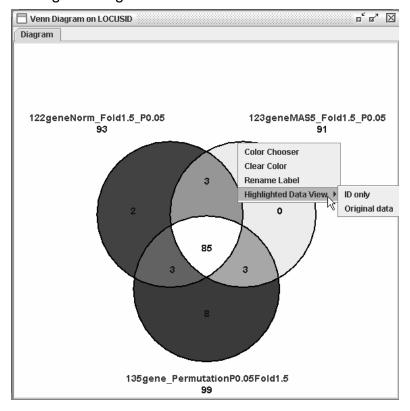
**Export** 

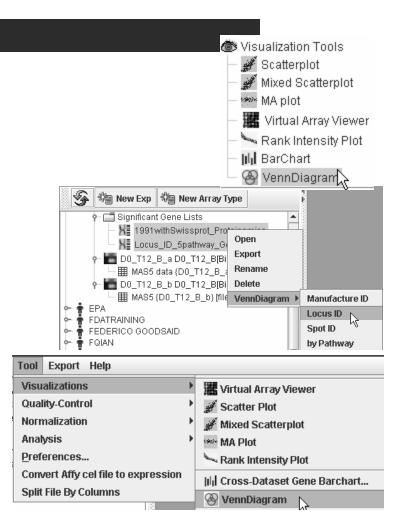
**Delete** 



## Significant Gene List(-cont)

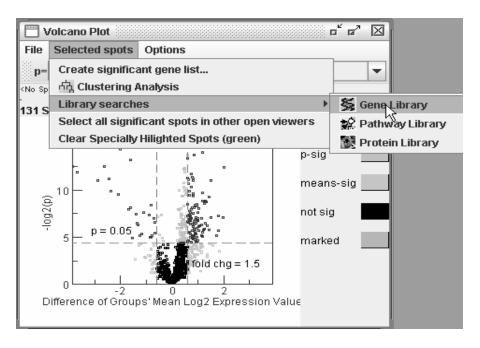
VennDiagram - get common genes from
 2~3 significant gene lists







#### Interpretation

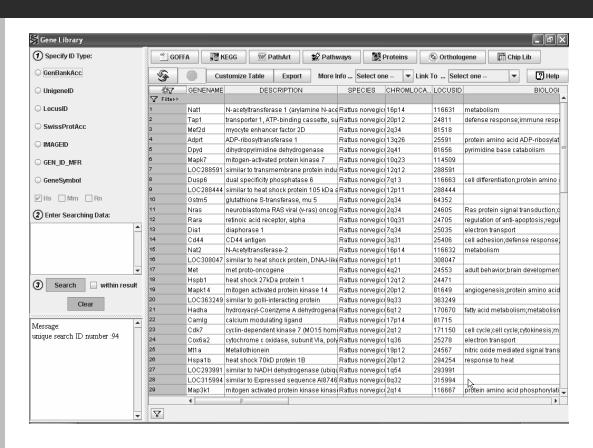


Link the significant genes to Gene Library for data interpretation





### Interpretation (continued)



- •The significant genes are listed here in Gene Library.
- Can search and sort the Gene library
- •There are links to other Libraries(Kegg, Pathart)





### Interpretation (continued)

- KEGG Kyoto Encyclopedia of Genes and Genomes <a href="http://www.genome.jp/kegg/">http://www.genome.jp/kegg/</a>
- KEGG is a suite of databases and associated software.
- KEGG Pathway database provides the information of metabolic, regulatory and disease pathways; Most of them are metabolic pathways.





### Interpretation (continued)

PathArt (Jubilant) – a pathway database

- •The Pathways (over 600 mammalian disease and signaling)
- •The Pathways is a collection of manually curated information from literature and public domain databases.

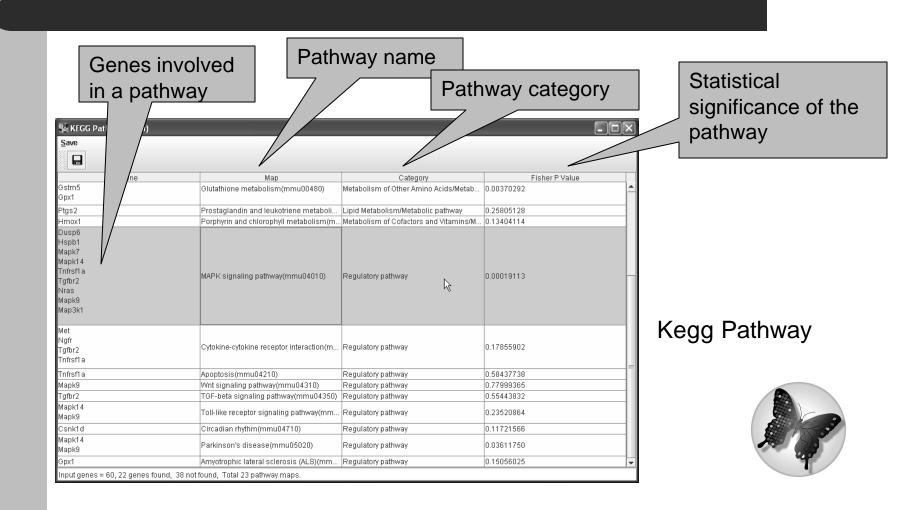
#### In ArrayTrack

	Human	Rat	Mouse
Kegg	134	116	124
PathArt	587	151	297





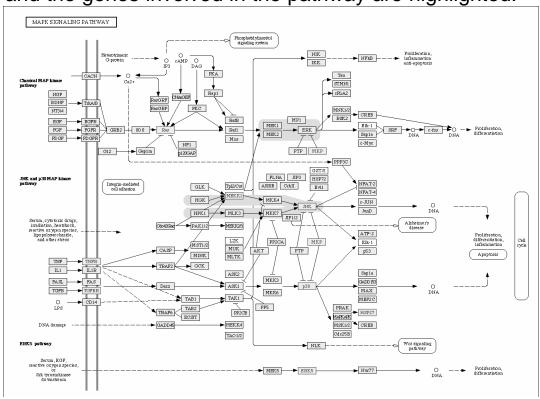
#### Interpretation (continued) Kegg





## Interpretation (continued) Kegg

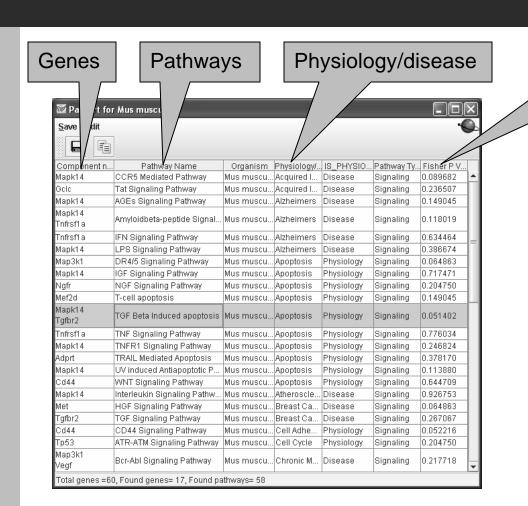
Double-click a specific pathway, the pathway map will be displayed and the genes involved in the pathway are highlighted.







#### Interpretation (continued) PathArt

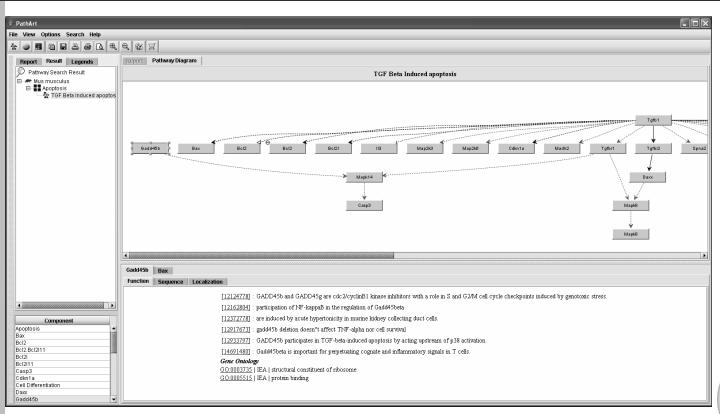


Statistical significance of the pathway





#### Interpretation (continued) PathArt







#### Interpretation (continued) GOFFA

- Developed based on Gene Ontology(GO) database
- Grouping the genes into functional classes
- GO- three ontologies
  - -Molecular function: activities performed by individual gene products at the molecular level, such as catalytic activity, transporter activity, binding.
  - -Biological process: broad biological goals accomplished by ordered assemblies of molecular functions, such as cell growth, signal transduction, metabolism.
  - -Cellular component: the place in the cell where a gene product is found, such as nucleus, ribosome, proteasome.



## Interpretation (continued) GOFFA

Select data type	Trong Trans and the state of th	NO NO	Gene Products
•	Tree Term Clustering All Genes P Path Plot	_ 1	NME6
○ GenBankAcc	□ □ all(49 P=1.000000)	2	CYP1A1
○ UnigenelD	🖟 🗂 biological_process(47 P=0.002730)	3	PTGS2
○ LocusID	□ □ development(14 P=0.004792)	4	NGFR
	□ ☐ morphogenesis(9 P=0.039353)	5	DAO1
○ SwissProtAcc	⊞ — ☐ organogenesis(8 P=0.045786)	6 7	TAP1 TNFRSF1A
GeneSymbol	tube morphogenesis(1 P=0.137460)	8	COX6A2
O Protein ID	😐 🗂 cellular morphogenesis(1 P=0.546747)	9	MET
O I TOKEM ID	■ 🗖 embryonic morphogenesis(2 P=0.054143)	10	VEGF
Select genome	⊕ 🗂 embryonic development(2 P=0.179509)		
O Demon S Mouse O Bot	□ □ reproduction(3 P=0.059708)		
○ Human ● Mouse ○ Rat	□ □ cell differentiation(1 P=0.794819)		
Input Data	□ □ □ growth(1 P=0.379243)		
OC288444	pigmentation(1 P=0.146151)		
DUC288444 DJB3	regulation of development(1 P=0.449401)		
MAPK14	mesoderm development(1 P=0.159028)		
RARA	physiological process(41 P=0.010023)		
HADHA	⊞ 🗂 behavior(2 P=0.114910)		
MEF2D	⊞ ☐ cellular process(23 P=0.582550)		
HMOX1	⊞ ☐ regulation of biological process(12 P=0.128288)		
MET	⊞ ☐ molecular function(46 P=0.122915)		
NAT2	□ □ cellular component(29 P=0.999926)		
NGFR	□		
ADPRT	extracellular space(10 P=0.235583)		
_OC288727	Extracellular space(10 1 = 0.233383)		
LOC308047 BACH			
CYP2C13	□ □ organelle(10 P=0.991753)		
_OC293991	⊕ ☐ protein complex(7 P=0.304521)		
COX6A2	No.		
CAMLG		_	
PTGS2	Search inTree	7  4	
CRYAB	Find terms containing		
DNAJA1	Time terms containing		
MAP3K1	● GO term ○ Gene name or gene symbol ○ P Value		
HSPB1			
	Course Clear		
Search GO	Search Clear		
Search GO		<b>7</b> 1	



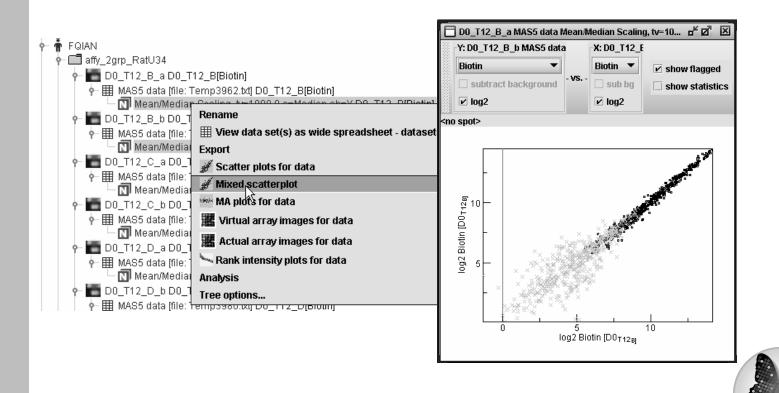


#### **Data Exploring**

- Scatter Plot
  - plot the fluorescence intensity data of Cy3 vs Cy5 for the same array
- Mixed Scatter Plot
  - compare two arrays in one plot, applies to both 2-channel and 1-channel data.
- Principal Component Analysis (PCA)
  - PCA is a way of identifying the data patterns and highlighting the data's similarity and difference.
- Bar Chart
  - displays expression data for a single gene across multiple arrays within the same experiment or across different experiment.
- HCA
  - is comprised of agglomerative methods and divisive methods that finds clusters of observations within a data set.

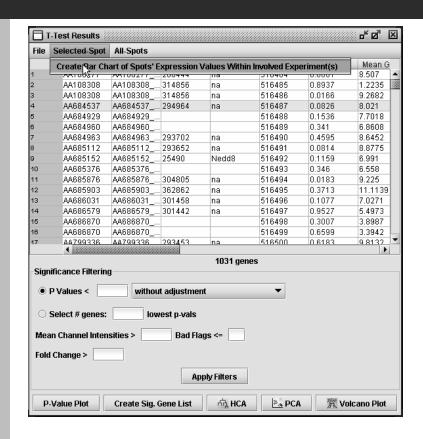


#### Data Exploring scatter plot





#### Data Exploring Bar chart



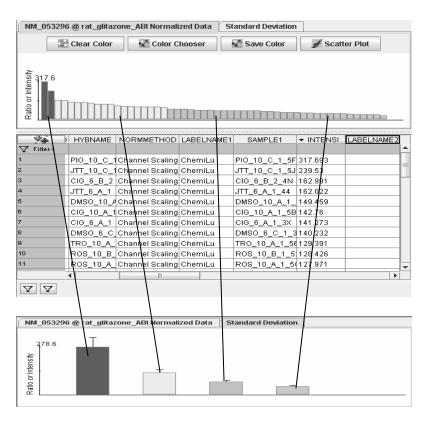
Access Bar chart from T-Test results.

Bar chart – display expression data for a single gene across multiple arrays in the same experiment or across different experiments.





#### Data Exploring Bar chart continued

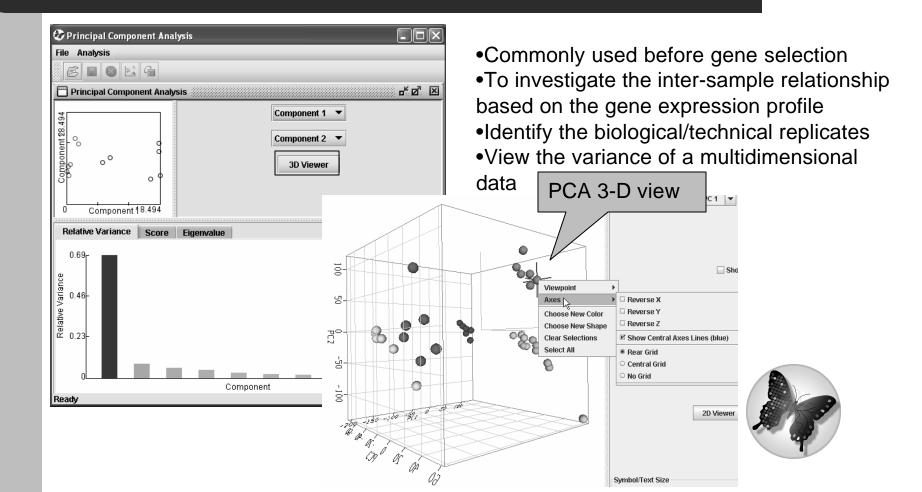


Grouping multiple arrays marked in different colors

Standard deviation bar chart for the above groups. The bar height represents the mean intensity, while the T-line above the bar stands for the value of SD. The color for each bar echoes the color of the bar chart at the top, in the same order.

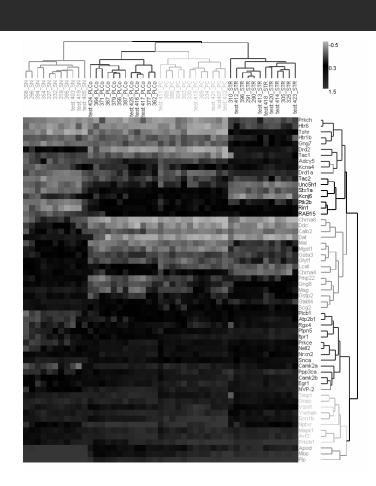


#### Data Exploring PCA





## Data Exploring HCA







#### **Accessing ArrayTrack**

 FDA Internal: <u>http://weblaunch.nctr.fda.gov/jnlp/arraytrack/index.html</u>
 <u>http://weblaunch.nctr.fda.gov/jnlp/arraytrack/citrix/index.html</u>

 FDA External: <a href="http://edkb.fda.gov/webstart/arraytrack/index.html">http://edkb.fda.gov/webstart/arraytrack/index.html</a>





#### **Technical Support**

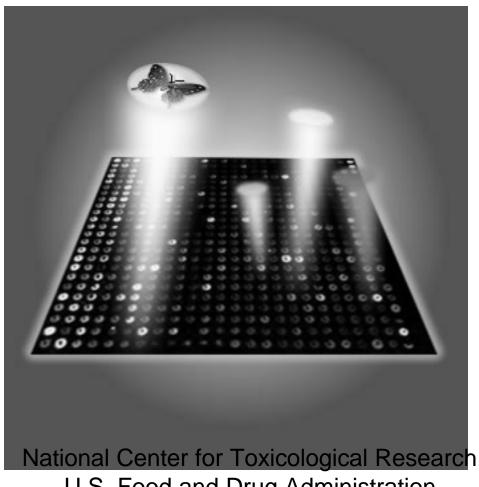
#### NCTRBioinformaticsSupport@nctr.fda.gov

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# Thank you!



U.S. Food and Drug Administration